

Chapter 19

The Role of Environmental Obesogens in the Obesity Epidemic

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19.1 Introduction

Nearly 260 years ago, prenatal exposure to a particular toxin, ethanol, was observed to have deleterious effects to the developing fetus [1]. During Prohibition in the United States, behavioral problems exhibited by children of “immoderate” alcohol-consuming families were attributed to poor social upbringing [2]. By the 1970s, methodical investigation of infants born to alcoholic mothers firmly established a prenatal component contributing to the common display of behavioral and mental problems [3, 4]. Acceptance of fetal alcohol syndrome (FAS) was slow because alcoholics were considered to be morally corrupt, social deviants who bred the same type of people [2]. Today, however, FAS is considered a rather obvious illustration of developmental programming because the teratogenic effects of ethanol are often robust and quickly discernable. Unfortunately, the effects of environmental chemicals that contribute to obesity and metabolic disease are not quite as visible, especially early in life. Major consequences of obesity such as diabetes and heart disease occur much later in life, and even early childhood obesity is more conveniently ascribed to a general lack of dietary restraint and/or lack of exercise. Indeed, the same stigma that plagued the acceptance of FAS is likely to affect the recognition and public awareness of the “obesity before birth” argument.

The conventional wisdom is that people can budget their diets as they do their finances: calories eaten should not exceed calories expended, otherwise, the body will drift toward a positive energy balance [5]. Suppose, though, that metabolic set points are perturbed from the beginning of life. For example, babies born to mothers who smoke exhibit a classic trend: low birth weight, with increased risk of obesity and metabolic syndrome later in life [6]. This pattern is characterized in the Developmental Origins of Health and Disease (DOHaD) hypothesis, which was developed after epidemiologists noted that poor maternal nutrition was correlated

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with reduced perinatal growth followed by a deliberate progression of adult disease in offspring [7–9]. In simple terms, the fetus “anticipates” that it will be born into a nutrient-deprived environment and makes suitable adjustments to its metabolic program. If the environment turns out to be food rich, then the adult will not be able to cope [10]. Low birth weight can be the outcome of multiple environmental stressors (not all nutrient based) and is indicative of a trajectory toward cardiovascular disease, diabetes, and obesity [11–14]. During certain critical developmental time windows (e.g., periconception), with relatively low exposure to a stressor, the fetus might appear completely normal [15], yet subtle developmental cues still exist and will manifest themselves later in life. Most studies supporting the DOHaD hypothesis are primarily nutritional in nature, thus the potential of environmental toxins to modulate developmental pathways that control energy and metabolism is mostly unexplored. Whether the stressor is coming from the outside, or produced endogenously by the parent, virtually all identified compounds that induce obesity are endocrine disrupting chemicals (EDCs). Many are either direct ligands for hormone receptors or affect components in metabolic signaling pathways under hormonal control.

19.2 An Archetype for Prenatal Chemical Exposure Leading to Obesity (Obesogens): Tributyltin

“Obesogens” are chemical compounds that can promote obesity by increasing the number of fat cells (and fat storage into existing fat cells), by changing the amount of calories burned at rest, by altering energy balance to favor storage of calories, and by altering the mechanisms through which the body regulates appetite and satiety.

Organotins are a family of compounds that contain at least one Sn—C bond. Invented in the 1850s, these “organic bodies of tin” were less a discovery of the compounds themselves than an educational instrument that encouraged the development of organometallic chemistry and the concept of valence number [16]. Currently, organotins are prevalent in industry, used in fungicides, wood preservatives, and heat stabilizers in polyolefin plastics [17]. Organotins were once widely used as antifouling paints on boats in the 1960–1970s, and have since been regulated, but not completely phased out [18]. Because organotins are lipophilic, they readily bioaccumulate in bacteria, algae, and aquatic invertebrates [19]. Concern over the biological consequences of organotin exposure was first reported when the female gastropod mollusk, exposed to tributyltin (TBT), developed a sperm duct, seminal vesicle, and penis [20, 21]. Neurotoxic [22] and mitochondrial toxic [23, 24] effects were also observed. Reproductive effects were first observed in higher vertebrates when it was noted that exposure to TBT masculinized genetically female Japanese flounder [25]. Subsequently, it was found that *Xenopus laevis* tadpoles exposed to low levels of tributyltin exhibited ectopic fat cell production; a novel and unexpected finding [26]. TBT was thus identified as the first “obesogen.”

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91 In mice, a single, prenatal exposure to TBT during gestation, results in prema-
92 ture accumulation of fat in adipose tissues [26] (Fig. 19.1). In accord with nutritional
93 experiments supporting the DOHaD hypothesis, the mice were born slightly under-
94 weight, but had already stored fat at the expense of total body mass [26]. By
95 6 months, the mice were significantl heavier, despite normal diet and exercise
96 (unpublished). Histological sections of newborn liver, testis, mammary gland, and
97 inguinal adipose tissue (which normally do not store lipids before feeding com-
98 mences) all showed pronounced lipid accumulation in the pups born to TBT-treated
99 mothers [26]. Hence, the tendency to store excess fat was already programmed
100 before the mouse was born, simply due to a single exposure of TBT, at a dose equiv-
101 alent to what humans might acquire, inadvertently, from their environment. TBT is
102 one of few examples of an exogenous chemical that generates an obese phenotype in
103 both sexes, solely due to in utero exposure. The mechanisms underlying why early
104 life exposure to chemicals, like TBT, might increase a person's propensity toward
105 obesity is the focus of this review.

19.3 Obesogens Acting on Estrogen/Androgen Metabolism

110 Estrogens in the adult are protective against android (abdominal) obesity and
111 metabolic disease. The ovariectomized rat (a model for menopause in women and
112 estrogen deficiency in men) has abdominal obesity, which is reversed upon treatment
113 with estrogen [27, 28]. In 3T3-L1 preadipocytes constitutively expressing the estrogen
114 receptor, triglyceride levels, and lipoprotein lipase expression are decreased
115 [29]. Conversely, knockout of the estrogen receptor leads to increased white adipose
116 depot size, central weight gain, and impaired glucose metabolism [30, 31]. The
117 same phenotypes seen with estrogen insufficiency can also be simulated by the inhibi-
118 tion of aromatase, an enzyme that converts testosterone to estradiol. Aromatase
119 knockout mice are obese and this phenotype is attributed to an estrogen deficit
120 as opposed to testosterone excess [32]. Metabolic disease, fatty liver, and abdomi-
121 nal obesity have also been demonstrated in a human case study where CYP19A1,
122 the gene encoding aromatase, is rendered non-functional [33]. Interestingly, trib-
123 utyltin (TBT) is an inhibitor of P450-mediated aromatase expression and/or action,
124 in both fish and humans [34, 35]. The masculinization of female mollusks by TBT is
125 thought to result from inhibition of aromatase; although there is also evidence that
126 TBT acts through nuclear receptors to inhibit aromatase expression. Whether and
127 to what degree TBT perturbs estrogen availability in mammals to affect adiposity is
128 currently unknown.

129 Prenatal exposure to excess estrogen does not protect the fetus against obesity,
130 despite the fact that estrogen promotes leanness in adults. The most prevalent model
131 for studying prenatal estrogen exposure is the diethylstilbestrol (DES) mouse. DES
132 is a synthetic estrogen that actually binds to the estrogen receptor two to three times
133 stronger than does the natural ligand, 17 β -estradiol [36]. When mice are treated
134 prenatally with low-dose DES, they give birth to pups that are initially smaller, but
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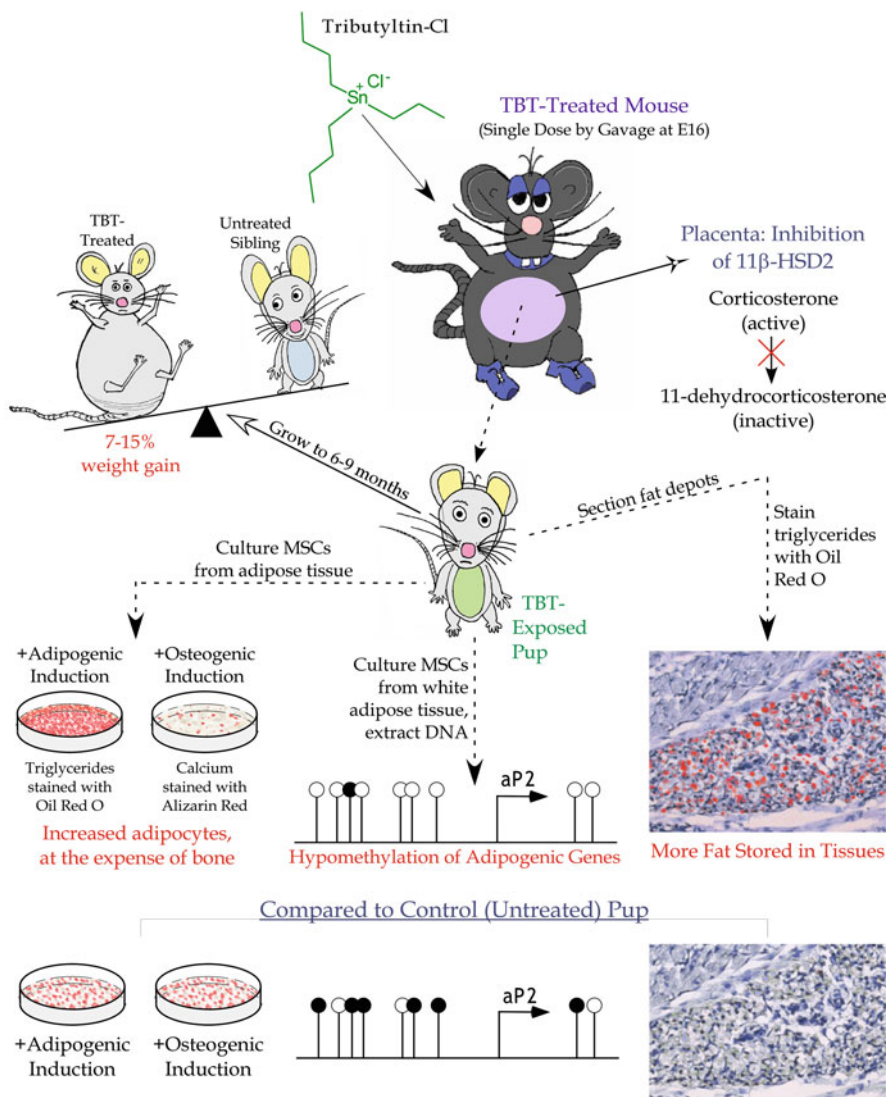


Fig. 19.1 Prenatal exposure to tributyltin (TBT) predisposes an organism to be obese. A single exposure to TBT at embryonic day 16 causes a host of changes in developing animals. TBT exposure alters the expression of 11β-HSD2 in the placenta, which could increase glucocorticoid levels. Prenatal TBT exposure produces pups that have already stored fat in tissues at birth, compared with control animals that do not. Multipotent stromal cells from exposed animals are predisposed to develop into adipocytes at the expense of bone cells. Promoters of some PPARγ target genes are undermethylated in MSCs from exposed animals compared to controls. This increased number of adipocytes may be important in the 7–15% weight gain seen in exposed animals from 6 to 9 months of age

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181 become heavier later in life [37]. The environmental estrogen bisphenol-A (BPA),
182 activates the estrogen receptor and is also an inverse antagonist (i.e., activator) of the
183 estrogen-related receptor [38]. BPA treatment of pregnant dams results in smaller
184 offspring that exhibit “catch-up” growth and are significantly heavier by 6 weeks
185 of age [39]. Dichlorodiphenyl-dichloroethylene (DDE), the major metabolite of the
186 pesticide DDT, is both an estrogen receptor activator and an anti-androgen [40, 41].
187 Mothers who lived along the Lake Michigan shoreline, exposed to high levels of
188 DDT, were more likely to have a child that exhibited elevated BMI in adulthood
189 [42]. One area of active study concerning the obesogenic effects of estrogen is the
190 regulation of leptin and its receptor by estrogens [43]. High levels of maternal leptin
191 are correlated with high fetal leptin [44] which is a precursor to weight gain later
192 in life. The obesogenic effects of estrogenic chemicals are explored more fully in
193 [Chapter 17](#).

19.4 Obesogens Acting on Glucocorticoid Metabolism

199 Disruption of glucocorticoid homeostasis also contributes to the pathogenesis of
200 obesity and metabolic syndrome. Obesity is linked to a general increase of positive
201 feedback within the hypothalamic–pituitary–adrenocortical (HPA) axis, leading to
202 an oversecretion of cortisol from the adrenal gland [45–49]. This does not neces-
203 sarily translate to higher circulating cortisol levels in the blood; and for that reason,
204 obesity is not associated with systemic hypercortisolism, as in the case of Cushing’s
205 syndrome. Rather, the hypercortisolism observed in obesity is intracellular and
206 peripheral in nature, often characterized by the impaired ability to clear or inactivate
207 cortisol in adipose tissue, particularly visceral adipose tissue [50]. Visceral adiposity
208 is particularly dangerous since it is associated with all the major health conse-
209 quences and mortality risk of obesity (reviewed in [51]). In addition, the visceral
210 adipose tissue itself is populated with more glucocorticoid receptors (GRs) than
211 subcutaneous tissue [52, 53]. Since glucocorticoids increase adipocyte proliferation
212 and their differentiation from stromal cells, the presence of excess glucocorticoids
213 or GRs will undoubtedly stimulate adipogenesis locally [54–56].

214 One popular hypothesis that links obesity to the HPA axis is the
215 dysregulation of 11 β -hydroxysteroid dehydrogenase type-1 (11 β HSD1) (see
216 [Chapter 13](#)). 11 β HSD1 is expressed wherever the glucocorticoid receptor is
217 found (i.e., ubiquitously) and primarily functions to convert the inactive cortisone
218 (or 11-dehydrocorticosterone in rodents) into the active cortisol (or corticosterone in
219 rodents) (reviewed in [57]). In humans, obesity and metabolic syndrome are asso-
220 ciated with elevated 11 β HSD1 [50, 58, 59]. The phenomenon is also observed in
221 obese Zucker rats, where 11 β HSD1 is increased specifically in omental adipose tis-
222 sue [60]. Concomitantly, the Zucker rats showed decreased 11 β HSD1 expression
223 in the liver, which might lead to a compensatory effect whereby the HPA is stimu-
224 lated to release more corticosterone [50]. The alternate dehydrogenase (11 β HSD2)
225 functions in the opposite direction; it converts active cortisol (or corticosterone)

226 to inactive cortisone (or 11-dehydrocorticosterone) and is mostly expressed in the
227 kidney, colon, salivary glands, and placenta, where mineralocorticoid receptors are
228 present [61–63]. 11 β HSD2 is expressed at low levels in adipose tissue and its reduction
229 might be correlated with obesity [64, 65]. However, deficiency of 11 β HSD2
230 at the kidney results primarily in increased mineralocorticoid receptor activation,
231 hypokalemia, and hypertension [66].

232 Excess glucocorticoid exposure during pregnancy is commonly associated with
233 lower birth weights, but increased risk of cardiovascular disease, diabetes, and
234 hypertension in the adult offspring (reviewed in [67]). Endogenous stimulation of
235 glucocorticoid levels in the mother can potentially wreak havoc on fetal metabolism.
236 In studies of maternal distress, where the mother's physiological state is negative
237 or depressed, the mother produces more corticotropin-releasing hormone, which in
238 turn leads to increased cortisol secretion, and reduced birth weight in the offspring
239 (reviewed in [68]). Exogenous stimulation of glucocorticoid levels in the mother is
240 also common. For example, ethanol exposure in guinea pigs led to elevated levels of
241 cortisol in the mother (measured in the saliva) which was transmitted to fetal blood
242 and amniotic fluid [69]. When pregnant rats were given the synthetic glucocorti-
243 coid dexamethasone, the birth weight of the offspring was reduced, followed by an
244 increased risk for hypertension in adults [70]. Monkeys treated with dexamethasone
245 during pregnancy had offspring that were normal at birth, but exhibited significant
246 weight gain at 2 months of age [71]. These monkey infants grew to develop obesity
247 and demonstrated all signs of metabolic syndrome (increased blood pressure, high
248 total cholesterol, decreased HDL, and insulin resistance).

249 Recently, a study showed that increased cortisol production rates in adults were
250 associated with visceral adiposity. Importantly, however, weight loss did not return
251 cortisol production to normal [72]. Obesity was not the source of the elevated corti-
252 sol production rates; rather, a perturbed HPA axis was a predisposing mechanism for
253 obesity. This mechanism might account in part for why people cannot lose weight
254 effectively. In this view, any environmental toxin that perturbs the HPA axis in early
255 life could also contribute to obesity and subsequent resistance to weight loss. There
256 are many possible mechanisms by which xenobiotic chemicals can influence gluco-
257 corticoid metabolism to disrupt energy balance, appetite, and the stress response
258 (reviewed in [73]). Given that 11 β HSD1 catalyzes the conversion of inactive to
259 active glucocorticoids, one possible way to disturb the HPA axis would be to render
260 11 β HSD1 hyperactive. Adult rats given a daily dose of alcohol throughout the first
261 22 days of gestation gave birth to offspring that demonstrated a significant increase
262 in 11 β HSD1 activity, with a subsequent increase in serum cortisol, in both liver and
263 adipose tissue [74]. Hence, it is no surprise that alcohol exposure in utero leads to
264 an activated HPA axis [75] and is associated with glucose intolerance in the adult
265 [76, 77].

266 Of course, the fetus is normally shielded against excess glucocorticoids because
267 placental 11 β HSD2 is highly expressed throughout pregnancy to reduce fetal
268 cortisol exposure [78]. Prenatal inhibition of 11 β HSD2 by carbenoxolone admin-
269 istration throughout pregnancy leads to reduced birth weight, anxious behavior,
270 and increased secretion of corticotropin-releasing hormone in rats [79]. Cadmium,

dithiocarbamates, and organotins all have the potential to reduce 11 β HSD2 activity in the placenta [80–82]. Hence, one plausible mechanism for obesity in offspring from mothers exposed to tributyltin (TBT) is that placental 11 β HSD2 activity is reduced. In fact, TBT is readily transferred from the mother and accumulates in the fetal placenta, liver, and brain [83]. Hence, the leaking of glucocorticoids to the fetus through the activation of 11 β HSD1, or placental inhibition of 11 β HSD2, is a potential mode of action for environmental chemicals. This is an attractive idea because the dehydrogenases function in the direct activation or inactivation of cortisol. However, due to the complexity of the HPA axis, there are many more avenues of endocrine disruption on glucocorticoid metabolism yet to be uncovered. For example, corticosteroid-binding globulin (CBG) activity is negatively correlated with BMI, waist-to-hip ratio, and insulin resistance in healthy adults [84]. Adipose tissue that is deficient in CBG cannot evacuate cortisol to the blood. In humans with CBG deficiency, preadipocytes readily proliferate and differentiate into adipocytes [85]. Hence, an EDC that lessened CBG activity might also lead to obesity in the adult.

19.5 Obesogens Acting on Peroxisome Proliferator-Activated Receptors

Unlike steroid receptors, which have long been associated with growth, metabolism, and reproductive development, the peroxisome proliferator-activated receptors (PPARs) were originally recognized because certain chemicals activated these receptors to increase peroxisome proliferation in rats [86, 87]. Recent research has focused on PPAR involvement in lipid metabolism and adipogenesis. PPAR α is primarily expressed in the liver and is stimulated upon starvation [88], where free fatty acids, once liberated from adipose tissue and transported into the blood, are oxidized in the liver, resulting in ketone bodies that provide energy to muscular tissue and the brain (reviewed in [89]). Fatty acids are natural ligands for PPAR α , although drugs such as fenofibrat and gemfibrozil (used to lower LDL levels) also increase PPAR α activity [90]. PPAR γ is primarily expressed in adipocytes [91] and upon activation, increases adipogenesis [92]. Thiazolidinediones (TZDs), which combat type 2 diabetes, are potent activators of PPAR γ [93]. PPAR α activation is thought to protect against obesity, whereas stimulation of PPAR γ is obesogenic. Therefore, PPAR γ has become a focus of many recent obesity-related studies. Nevertheless, chronic stimulation of PPAR α leads to glucose intolerance, as well as deterioration of the heart muscle due to lipotoxicity [94–96]. Thus, while PPAR α converts fat into energy, it can limit the uptake of glucose into tissues, leading to diabetes.

Like all PPARs, PPAR γ forms a heterodimer with the retinoid-X receptor (RXR), a binding partner for multiple nuclear receptors. The ligand-binding pocket of PPAR γ is large [97] and can accommodate various chemical structures [98]. PPAR γ promiscuity is well demonstrated by the fact that tributyltin (TBT) has nanomolar affinity for PPAR γ , yet its structure is highly dissimilar to fatty acids and TZDs

316 [26, 99]. Indeed, the mechanistic basis for TBT-promoted adipogenesis is most
317 strongly supported by evidence that TBT is an agonist for PPAR γ and RXR [100].
318 Competitive binding assays show that TBT has comparable affinity for RXR, as do
319 synthetic RXR agonists [26]. TBT was actually a better activator of PPAR γ even
320 compared to TZDs such as troglitazone [26, 100]. The crystal structure of TBT
321 along with the RXR α ligand-binding domain plus a coactivator fragment shows that
322 TBT binds covalently to RXR [101], which means that it will not readily dissociate
323 once attached. While there is no structural evidence for TBT association with
324 PPAR γ , the PPAR γ antagonist, T0070907 [102], reduced TBT-stimulated adipogenesis,
325 indicating that the obesogenic activity of TBT is likely mediated through
326 activation of PPAR γ [103].

327 Inappropriate stimulation of PPARs by EDCs is a potential factor underlying
328 the etiology of many forms of obesity. This point is perhaps best made by pharmaceutical
329 obesogens (TZDs) used to treat diabetes. Because TZDs are effective
330 agonists of PPAR γ , they promote weight gain in humans by increasing adipose tissue
331 differentiation [104, 105], even as they improve insulin resistance and blood
332 glucose. Since drugs aberrantly modulate PPAR γ to stimulate adipogenesis, EDCs
333 like TBT can do the same. However, TBT is not the most ubiquitous EDC to which
334 humans are exposed. Some phthalates (see Chapter 16) are also PPAR γ agonists
335 [106] and stimulate the proliferation of adipocytes in the 3T3-L1 cell culture model
336 [107]. Phthalate metabolites are associated with increased waist circumference in
337 men [108], and therefore, are predicted to be obesogenic. Phthalates are ubiquitous
338 organic chemicals that give plastics such as polyvinyl chloride (PVC) with more
339 flexibility and durability. Since phthalates are not permanently bonded to PVC, they
340 readily leach into food, from medical devices and materials used in construction
341 and manufacturing. Phthalates are probably able to cross the fetoplacental barrier,
342 as demonstrated by a simulated human placental perfusion *ex vivo* model [109].
343 Identifying the role of phthalates as obesogens is an important topic for future
344 research.

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19.6 Epigenetics: The Gene–Environment Connection

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350 Obesogens are predicted to influence the prenatal programming of obesity through
351 epigenetic modification of gene activity; an attractive mechanism frequently
352 invoked to explain complex environment–gene interactions [110]. There are more
353 than 6,000 genes expected to influence body weight. Of these, about 10 times more
354 genes favor the increase rather than decrease of weight [111]. Epigenetic modification
355 of any of these genes theoretically could result in “developmental plasticity,”
356 which allows an organism to make rapid adaptations to changing environments by
357 altering levels of gene expression via DNA methylation, modification of histone proteins,
358 or alteration of mitochondrial function [112–114]. During development, the
359 embryo undergoes genome-wide DNA demethylation and remethylation [115–117],
360 which is coupled to corresponding changes in histone acetyltransferase activity

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361 [118]. The first wave of methylation reprogramming occurs prior to the blastocyst
362 stage and establishes which cells will commit to a particular lineage [119]. The
363 second wave occurs during the development of the testes or ovaries, where the pri-
364 mordial germ cells incorporate sex-specific imprinting patterns [120]. During both
365 of these time windows, an exogenous chemical can impose changes (in some cases
366 permanent) in the methylation status of DNA.

367 Alterations in the methylation status of certain genes have been observed in off-
368 spring that experience malnutrition in the womb. Fetal liver derived from rats fed a
369 low-protein diet showed promoter hypermethylation in the liver X-receptor (LXR)
370 [121] and hypomethylation in PPAR α [122]. The methylation-deficient status of
371 PPAR α was rescued by supplementing the low-protein diet with folic acid, a methyl
372 donor [122]. These modifications could plausibly disturb normal lipid homeostasis
373 and impair metabolism, perhaps leading to obesity. Chemical-induced alterations in
374 DNA methylation status have also been observed for diethylstilbestrol (DES) [123]
375 and the dioxin TCDD [124]. In preimplantation embryos, DNA methyltransferase
376 activity was altered depending on whether the embryo was exposed to TCDD, DES,
377 or polychlorinated biphenyl-153 (PCB153) [125]. Liver nuclear extracts derived
378 from TBT-treated rats showed increased histone acetyl transferase activity [126].
379 Therefore, by affecting the levels of enzymes, such as DNA methyltransferase and
380 histone acetyl transferase, which have broad impacts on gene regulation, these EDCs
381 can conceivably transmogrify the entire epigenetic landscape. Hence, exposure to
382 environmental chemicals has the potential to remodel the behavior of the myriad of
383 genes that are connected to obesity. **should read "wherein"**

384 The maternal programming of adipose tissue development is fundamentally con-
385 cerned with epigenetic modification that alter stem cell fate. Adipogenesis is a
386 differentiation event in the mesodermal lineage where in multipotent stromal cells
387 (MSCs) or more restricted derivatives give rise to fat cells. MSCs have the potential
388 to mature into various tissues including bone, muscle, cartilage, or adipose [127].
389 MSCs harvested from epididymal or ovarian fat pads of mice exposed to TBT in
390 utero differentiate into significantly more fat cells, compared to controls [103].
391 Indeed, adipogenesis occurred at the expense of osteogenesis, suggesting that pre-
392 natal exposure to TBT biased the MSC population toward the adipogenic lineage
393 [103]. TBT likely induces epigenetic changes within the MSC compartment that
394 promote demethylation of adipogenic genes. It is currently controversial whether
395 active demethylation of adipogenic genes (with a corresponding up-regulation in
396 expression) is indicative of MSC differentiation into adipocytes. Recent evidence
397 shows that in a population of pure adipose-derived MSCs, adipogenic genes are
398 already hypomethylated in the MSCs, suggesting that MSCs are predisposed to
399 adipogenesis [128]. Hence, an environmental chemical might augment the innate
400 hypomethylated state of adipogenic genes or increase the methylation of other lin-
401 eages, such that the MSC pool is biased prior to adipocyte differentiation. In support
402 of this model, uninduced MSCs harvested from mice exposed to TBT in utero
403 show decreased methylation in the promoter region of fatty acid-binding protein
404 4 (FABP4), a marker of adipocytes; suggesting that the MSC population has already
405 been epigenetically modified to favor adipogenesis [103]. Future experiments will

406 be required to identify what other genes are required to mediate the effects of
407 prenatal TBT exposure in the MSC compartment, committing more MSCs to the
408 adipogenic lineage.

409 About 40 years ago, in an effort to properly measure and define obesity,
410 researchers postulated that the number of fat cells in the adult is fixed and that people
411 gain or lose weight by filling or emptying these cells with fat [129, 130]. Recently,
412 this hypothesis was proven correct, but with an important modification. The number
413 of fat cells in an adult is essentially constant as had been supposed. However, in
414 contrast to previous dogma, adipocytes are continually undergoing apoptosis [131]
415 and being replenished [132]. Using the same [^{14}C]-labeling method employed to
416 prove that adults have the capacity to produce new brain cells, researchers have
417 shown that the life span of a fat cell is approximately 10 years in both obese and
418 lean subjects [133]. During severe weight loss (e.g., gastric bypass), the number of
419 fat cells remained constant, while cell volume decreased [133]. This suggests that
420 obese individuals possess a pool of MSCs that is intrinsically biased toward replen-
421 ishing fat cells, i.e., that obese individuals have a steady state level of adipocytes that
422 may be higher than non-obese people. Such a bias could be regulated by epigenetic
423 changes due to exposure to environmental cues experienced during critical devel-
424 opmental windows. Chemicals like TBT influence the number of MSCs committed
425 toward the adipogenic fate by remodeling the chromatin, favoring the expression of
426 genes that promote adipogenesis. Notwithstanding these recent advances, there is
427 much yet to be learned about how epigenetic contributions modulate the prenatal
428 programming of MSC fate and what role obesogens play in this process.

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432 19.7 Conclusion

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Prenatal exposure to obesogens is likely to be an underestimated contributor to the obesity epidemic. Although unhealthy food consumed in large portions together with insufficient physical activity are likely to be among the chief substrates of weight gain, rates of obesity have increased in infants [134], as well as children and adults. This suggests that obesity is being programmed prenatally or in early childhood. There is increasing evidence that supports the proposal that environmental chemicals may contribute to the prenatal programming of obesity (see [Chapter 16](#)). Prenatal exposure to tributyltin, a chemical for which the mechanism of action is known, predisposes organisms to obesity, suggesting that the DOHaD model is applicable to effects of chemical exposure, in addition to altered nutrition. There are numerous EDCs more prevalent in the environment than tributyltin that have been linked to metabolic disease and whose increased usage mirrors the rising trend of obesity. The metabolic pathways targeted by most of these chemicals remain to be determined and firm links between chemical exposure and obesity should be based on understanding the underlying mechanisms. Understanding how chemicals enter the body and are transferred to the developing fetus is still not well understood. Epigenetics and the regulation of the stem cell lineages will

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451 provide answers to the mechanistic questions regarding how obesogens disrupt the
 452 endocrine system. This will inform the development of therapeutics and perhaps
 453 even non-pharmacologic solutions to the obesity problem.

454 Lastly, an important policy issue raised by our work is that of personal respon-
 455 sibility. While it is undeniable that the balance of our food intake and activity is
 456 reflecte in total body weight, it is equally true that obesity is a multifactorial
 457 disease with inputs from many different developmental pathways. Works in our
 458 laboratory and elsewhere have revealed critical roles for EDCs in body weight. A
 459 more complete understanding of how these pathways impact the body's homeostatic
 460 mechanisms for energy balance, adipocyte number, appetite, and satiety will allow
 461 future research to move forward without being limited by the simplistic model that
 462 caloric intake and exercise can be trivially balanced like a checkbook to achieve
 463 optimum weight.

464 **Acknowledgments** Work in the authors' laboratory was supported by a grant from the NIH R01
 465 ES015849. A.J. is a pre-doctoral trainee of NSF IGERT DGE 0549479.

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Chapter 19

| Q. No. | Query |
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| AQ1 | "A. Janesick" has been set as corresponding author. Please check. |
| AQ2 | Please provide complete details for reference "18". |
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B. Blumberg should be listed as corresponding author

AQ2: Reference 18 is a web site <http://nsglc.olemiss.edu/Advisory/Antifouling.pdf>

AQ4: Reference 71 is an abstract from the 11th Annual Meeting of the Neuroendocrinology Section of the German Society of Endocrinology (DGE). It was published in the *Experimental and Clinical Endocrinology & Diabetes* (Volume 115, Issue 8). There is no page number given, but the abstract number is N14.